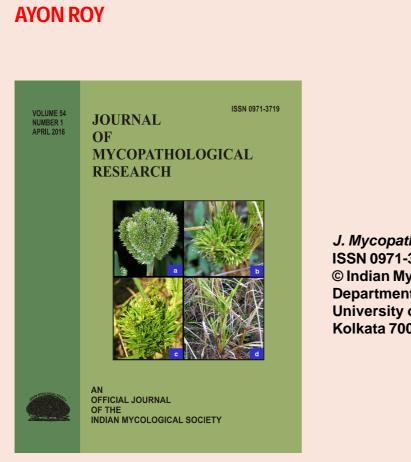
In vitro evaluation of some new fungicides against Blast and Sheath blight pathogens in rice



J. Mycopathol, Res, 54(1) : 71-75, 2016; ISSN 0971-3719 © Indian Mycological Society, Department of Botany, University of Calcutta, Kolkata 700 019, India

This article is protected by copyright and all other rights under the jurisdiction of the Indian Mycological Society. The copy is provided to the author(s) for internal noncommercial research and educational purposes.

In vitro evaluation of some new fungicides against Blast and Sheath blight pathogens in rice

AYON ROY

Department of Plant Pathology, Uttar Banga Krishi Viswavidyalaya, Pundibari 736165, Cooch Behar, West Bengal

Received : 12.06.2015

RMS Accepted : 17.07.2015

Published : 25.04.2016

Ten fungicides were evaluated in vitro for their potential against *Pyricularia oryzae* and *Rhizoctonia solani* causing blast and sheath blight in rice. Against P. oryzae, Tricyclazole 75WP and Azoxystrobin 18.2%+Difenoconazole 11.4% SC were found most effective resulting 84.31% and 80.00% reduction of mycelia growth at 100 ppm, respectively. Difenoconazole 25 EC was found effective at higher concentration (150 ppm) resulting 84.31% reduction of pathogen growth while Pencycuron 22.9SC was least effective against *P. oryzae*. EC50, an indicator of fungicide power against a particular fungus showed that Azoxystrobin 18.2%+ Difenoconazole 11.4% SC had highest toxicity against *P. oryzae* followed by Tricyclazole.75WP. Cyproconazole 50S and Azoxystrobin 18.2% + Difenoconazole 11.4%SC were found to be the most effective fungicides against *R. solani* resulting 100% reduction of pathogen growth at 150 ppm but Propiconazole 25EC was most toxic in term of its lowest EC50.

Key words: Fungicides, Pyricularia oryzae, Rhizoctonia solani, EC50

INTRODUCTION

Among the various fungal diseases of rice blast and sheath blight hold major attention during past few decades due to their recurrent appearance in all rice growing regions of the world. Rice blast caused by *Pyricularia oryzae*, an anamorph of *Magnaporthe grisea* attacks on stem nodes, leaves and all portions of the panicle and grains of rice (Seebold *et al*, 2004). Blast epidemic causes the complete defeat of seedling at the nursery and in field condition and accomplish 50-90% of total yield fatalities (Agrios, 2005). Sheath blight caused by *Rhizoctonia solani*, an anamorph of *Thanatephorus cucumeris* is another major production constraint in many rice growing areas especially in coastal and high humid regions. Loss in yield may upto 53% depending on the cultivar, environmental condition, stages at which the plants are infected and level of infection has been reported (Rajan, 1987; Roy, 1993). The disease is particularly important in intensive rice production systems (Savary and Mew, 1996). The natural infection of the sheath blight disease occurs at the seedling, tillering and booting stages of rice. The entire plant often gets killed under severe cases. The pathogen has a wide host range and can infect plants belonging to more than 32 plant families and 188 genera (Gangopadyay and Chakrabarti, 1982).

The most usual approaches for the management

of the rice fungal diseases include planting of resistant cultivars, application of fungicides, manipulation of planting times, fertilizers and irrigations (Georgopoulos and Ziogas, 1992; Moletti, 1988; Mbodi et al, 1987). The inherent level of resistance in rice to blast and sheath blight is very low. For farmers in developing countries, like Vietnam, India, Indonesia planting resistant varieties is the easiest and safest method but the resistance of most varieties is readily overcome within short period of time (Bonman et al, 1992). Chemical control of the diseases, therefore, is the most widely accepted strategies to keep the severity below economic threshold level but due to sudden mutation in certain population the resistance to fungicides may developed and the virulent population causes infection even on resistant varieties. Approximately 30 fungicides belonging to benzimidazoles, triazoles, succinate dehydrogenase inhibitors, melanin biosynthesis inhibitors, strobulirins, antifungal antibiotics, etc. have been tested and registered in India for control of blast and sheath blight (Kumar et al, 2014) and several new molecules are under testing. Yet the efficiency of particular fungicides could vary from place to place or from dosage to dosage. Farmers are advised to revolve the fungi-toxicants used to prevent the infectious fungus form rising resistance against those fungicides (Tangdiabang and Pakki, 2006). Hence, information about efficient fungicides with different modes of action should be offered to farmers. In this view, the present investigation was undertaken to assess the efficacy of some new commercially available fungicides against blast and sheath blight diseases.

MATERIALS AND METHODS

The test pathogens *P. oryzae* and *R. solani* were isolated from the infected rice leaf and sheath collected from Instructional Farm, Uttar Banga Krishi Viswavidyalaya, Pundibari, Cooch Behar, West Bengal. The pure cultures of the pathogens were maintained on Rice Extract Agar (REA) and Potato Dextrose Agar slants, respectively at $4\pm1^{\circ}$ C.

The comparative toxicity of ten fungicides on the growth of the pathogens under in vitro condition was evaluated by poisoned food technique (Dhingra and Sinclair, 1985). Fungicides like Propiconazole 25EC (Tilt), Tricyclazole 75WP (Baan), Tricyclazole 34.2% + Propiconazole 10.7% (Filia 52.5 SE), Difenoconazole 25EC (Score), Cyproconazole 50S

(Alto), Triamaphos 48EC (Kitazin), Carbendazim 50WP (Bavistin), Pencycuron 22.9SC (Monceren), Azoxystrobin 23SC (Amister) and Azoxystrobin 18.2%+Difenoconazole 11.4% SC (Amister Top 325 SC) at different concentrations (25, 50, 100 and 150 ppm) were used for in vitro assay. The fungicides were incorporated into the sterilized REA and PDA medium for the study on P. oryzae and R. solani, respectively. The sterilized petriplates containing amended media were inoculated with 6 mm disc of freshly prepared culture of the pathogens and incubated at 27±1°C for 7 days. Five replications for each concentration of an individual fungicide along with check by inoculating mycelia disc on non-amended media were maintained. The efficacy of°the fungicides was expressed as percent of radial growth over control, which was calculated by using the formula $I=(C-T/T)\times 100$ Where, I= Per cent inhibition over control C= Radial growth in control and T= Radial growth in treatments. The EC50 (i.e., the effective concentration for 50% inhibition in growth) of each fungicides against each pathogens was also calculated by working out the linear equation from the calculated per cent inhibition.

RESULTS AND DISCUSSION

Results on *in vitro* evaluation of the fungicides against P. oryzae have been presented in Table 1. It was observed that with increasing concentration of fungicides the mycelia growth of P. oryzae reduced accordingly. Maximum inhibition of 70.59-84.31% had been recorded against Tricyclazole 75WP at 25 ppm-100 ppm. The result was statistically at par with Azoxystrobin 18.2% + Difenoconazole 11.4%SC (69.02, 75.69 and 80.00% reduction at 25, 50 and 100 ppm, respectively). However, at 150 ppm Tricyclazole 75WP significantly outperformed (88.63% reduction in growth over control) Azoxystrobin 18.2% + Difenoconazole 11.4%SC (81.57% reduction in growth over control). Although at lower concentrations (25-50 ppm) Difenoconazole 25EC was moderately effective but at higher concentration it was good enough to reduce the mycelia growth of P. oryzae (77.65 and 84.31% reduction at 100 and 150 ppm, respectively). Pencycuron 22.9SC was the least effective showing 18.82% reduction in growth at 150 ppm concentration. Lowest EC50 was calculated for Azoxystrobin 18.2% + Difenoconazole 11.4%SC (1.50 ppm) closely followed by Tricyclazole 75WP (2.93 ppm). The other

Fungicide	Per cent reduction in growth over control				Linear equation	R ²	EC50
	25 ppm	50 ppm	100 ppm	150 ppm			(ppm)
Propiconazole 25%EC	41.96	43.14	47.45	56.86	y=17.58x+15.37	0.792	93.29
Tricyclazole 75WP	70.59	78.82	84.31	88.63	y=22.63x+39.42	0.993	2.93
Tricyclazole 34.2% + Propiconazole10.7%	51.37	60.78	67.84	74.12	y=28.44x+11.81	0.995	22.02
Difenoconazole 25%EC	49.80	66.27	77.65	84.31	y=43.88x-10.27	0.990	23.63
Cyproconazole 50%S	52.55	59.61	71.37	79.22	y=34.43x+3.07	0.982	23.07
Triamaphos 48%EC	25.10	38.43	49.02	53.73	y=36.93x-25.59	0.991	111.39
Carbendazim 50%WP	32.16	53.33	64.31	71.37	y=49.29x-34.34	0.973	51.42
Pencycuron 12.5%SC	0.78	6.67	12.55	18.82	y=22.42x-31.06	0.982	4125.93
Azoxystrobin 23SC	53.33	59.22	61.57	69.41	y=18.60x+27.05	0.916	17.13
Azoxystrobin 18.2%+Difenoconazole 11.4% SC	69.02	75.69	80.00	81.57	y=16.16x+47.16	0.973	1.50
SEm±	3.37	1.72	1.58	1.12			
CD(P=0.05)	10.08	5.14	4.73	3.34			

Table 1 : In vitro evaluation of fungicides against P. oryzae

test fungicides were in the rank of Azoxystrobin 23SC <Tricyclazole 34.2% + Propiconazole 10.7%SE < Cyproconazole 50S < Difenoconazole 25EC < Carbendazim 50WP< Propiconazole 25EC < Triamaphos 48EC whereas, Pencycuron 22.9SC showed highest EC50 (4125.93 ppm). Gohel *et al*, (2008) evaluated 19 fungicides against *M. oryzae* and found that Tricyclazole, Mancozeb, Carbendazim, Iprobenfos, Propiconazole and Edifenphos were highly effective against the test fungus. The results also corroborate earlier findings by Sood and Kapoor (1997), Ghaznafar *et al*, (2009), Jamal Uddin *et al*. (2011), Verma and Santhakumari (2012) and Singh *et al*, (2014).

The fungitoxicity of the test chemicals on R. solani revealed that as a general phenomenon with fungicide gradient the mycelial growth decreased significantly irrespective of fungicides concerned. Cyproconazole 50S and Azoxystrobin 18.2% + Difenoconazole 11.4%SC were found to be the most effective fungicides reducing the mycelia growth by 76.08-78.04% at 25 ppm to 100% at

150 ppm concentration. At 25, 50 and 100 ppm concentration, Propiconazole 25EC was statistically at par with Cyproconazole 50%S and Azoxystrobin 18.2% + Difenoconazole 11.4%SC exhibiting 73.73, 78.04 and 86.27% reduction in growth over control, respectively. Propiconazole 25EC at 150 ppm concentration resulted 90.98% reduction in mycelia growth of R. solani which statistically differed from the ability of Cyproconazole 50S and Azoxystrobin 18.2% + Difenoconazole 11.4%SC. Most of the other fungicides (Difenoconazole 25EC, Triamaphos 48EC, Carbendazim 50WP, Pencycuron 22.9SC and Azoxystrobin 25SC) at 100-150 ppm concentration resulted >50% growth inhibition. At lower concentration of 25ppm, 46.27% growth inhibition of R. solani was recorded with Carbendazim 50WP and was statistically at par with Triamaphos 48EC, Pencycuron 22.9SC and Azoxystrobin 23SC. Among all the fungicides, Tricyclazole 75WP showed least inhibition at all the concentrations evaluated. It was observed that Propiconazole 25EC had lowest EC50 (2.45 ppm) followed by Cyproconazole 50S and Azoxystrobin

: 54(1) April, 2016]

Table 2 : In vitro evaluation of fungicides against R. solani

Fungicide	Fungicide Per cent reduction in growth over control						EC50 (ppm)
	25 ppm	50 ppm	100 ppm	150 ppm			(ppin)
Propiconazole 25%EC	73.73	78.04	86.27	90.98	y=22.56x+41.23	0.980	2.45
Tricyclazole 75WP	7.84	34.12	47.06	52.55	y=56.79x-67.87	0.953	119.00
Tricyclazole 34.2% + Propiconazole10.7%	54.51	72.94	77.25	80.00	y=31.41x+14.05	0.877	13.95
Difenoconazole 25%EC	59.61	69.41	75.69	79.61	y=25.31x+25.05	0.987	9.68
Cyproconazole 50%S	78.04	80.78	95.29	100.00	y=30.30x+33.42	0.930	3.53
Triamaphos 48%EC	56.86	60.39	61.57	65.49	y=10.00x+42.88	0.928	5.15
Carbendazim 50%WP	46.27	52.55	60.00	63.14	y=22.10x+15.29	0.997	37.20
Pencycuron 12.5%SC	40.78	48.24	50.20	63.53	y=25.45x+4.39	0.847	61.96
Azoxystrobin 23SC	49.02	51.37	56.08	60.39	y=14.41x+28.01	0.952	33.58
Azoxystrobin 18.2%+Difenoconazole 11.4% S	SC 76.08	82.35	89.41	100.00	y=29.07x+34.08	0.942	3.53
SEm±	4.00	3.11	3.57	2.35			
CD(P=0.05)	11.98	9.30	10.71	7.03			

18.2% + Difenoconazole 11.4%SC (3.53 ppm). The other test fungicides were in the rank of Triamaphos 48EC < Difenoconazole 25EC < Tricyclazole 34.2% + Propiconazole 10.7% SE< Azoxystrobin 23SC < Carbendazim 50WP < Pencycuron 22.9SC < Tricyclazole 75WP. Jones et al. (1987) reported that Propiconazole at 100 ppm concentration resulted 96% reduction of mycelial growth of R. solani. The results of the present investigation were similar to the observations made by Biswas (2002), Chahal et al, (2003), Sunravadana et al, (2007), Bhuvaneswari and Raju (2012). Swamy et al (2009) screened some new fungicides against R. solani and found that Tricyclazole and Propiconazole had good impact on inhibition of pathogen growth. Haggag and El-Gamal (2012) studied the in vitro effect of Propamocarb, Hymexazol, Pencycuron, Flutolanil and Thiophanate methyl against R. solani and observed that Hymexazol significantly reduced the mycelia growth of the tested pathogenic fungal isolates, followed by Pencycuron, Propamocarb, Thiophanate methyl and Flutolanil. EC50 is used only to compare the power of fungicides in the control of a specific fungus by identifying the most efficient one having no relation with the actual rate used in field. Hence, present investigation extends the sphere of fungicide options for curative management of blast and sheath blight with triazoles like Cyproconazole 50S, Difenoconazole 25EC and some mixture fungicides like Azoxystrobin 18.2% + Difenoconazole 11.4%SC and Tricyclazole 34.2% + Propiconazole 10.7%SE.

ACKNOWLEDGEMENTS

The author is grateful to Syngenta India Limited for providing the financial assistance and the new fungicide formulations.

REFERENCES

Agrios, G.N. 2005. *Plant Pathology* Fifth Edition. Elsevier Academic Press. pp. 343.

- Bhuvaneshwari, V. and Raju, K.S. 2012 Efficacy of new combination fungicide against rice sheath blight caused by *Rhizoctonia solani* (Kuhn.). *J. Rice Res.* **5**: 57-61.
- Biswas, A. 2002, Evaluation of new fungicidal formulations for sheath

blight control. J. Mycopathol. Res. 40: 9-11.

- Bonman, J. M., Khush, G. S. and Nelson, R. J. 1992 Breeding rice for resistance to pests. Ann. Rev. Phytopathol, 30: 507-528.
- Chahal, K.K.S., Sokhi, S.S. and Rattan, G. S. 2003. Investigation on sheath blight of rice in Punjab. *Indian Phytopath.* 56: 22-26.
- Dhingra, O.D and Sinclair, J.B. 1985. *Basic Plant Pathology Methods*. 2nd Edition. CRC Lewis Publishers, London. 434 pp.
- Gangopadhyay, S. and Chakrabarti, N. (1982). Sheath blight of rice. *Rev. Pl. Path.* **61:** 451-460.
- Georgopoulos, S.G, and Ziogas, B.N, (1992). Principles and methods for control of plant diseases, Athens, 236 p.
- Ghazanfar. M.U., Waqas Wakil., S.T. Sahi and Saleem-il-Yasin. (2009). Influence of various fungicides on the management of rice blast disease, *Mycopath* 7: 29-34.
- Gohel N.M., Chauhan, H.L. and Mehta, A.N. 2008. Bio-efficacy of fungicides against *Pyricularia oryzae* the incitant of rice blast. J. Plant Dis. Sci. **3:** 189-192.
- Haggag, K.H.E. and El-Gamal, N.G. 2012. In vitro study on *Fusarium solani and Rhizoctonia solani* isolates causing the damping off and root rot diseases in tomatoes. *Nature and Science* **10**: 16-25.
- Jamal Uddin, Hajano, A., Mubeen, L., Mumtaz, A., Pathan, M., Ali, K. Shah, G.S. 2011. In vitro evaluation of fungicides, plant extracts and biocontrol agents against rice blast pathogen *Magnaporthe oryzae* Couch. *Pak. J. Bot.* 44: 1775-1778.
- Jones, R.K., Belmar, S.B. and Jeger, M.J. 1987. Evaluation of Benomyl and Propiconazole for controlling sheath blight of rice caused by *Rhizoctonia solani*. *Plant Dis.* **71**:222-225.
- Kumar. P.M.K., Sidde Gowda, D.K., Rishikant, M., Kiran Kumar, N., Pandurange Gowda, K.T. and Vishwanath, K. 2013 Impact of fungicides on rice production in India In: Fungicides – showcases of integrated plant disease management from around the world (http://dx.doi.org/10.5772/51009). pp. 77-98.
- Mbodi, Y., Gaye, S. and Diaw, S. 1987. The role of tricyclazole in

rice protection against blast and cultivar improvement. *Parasitica*, **43:** 187-198.

- Moletti, M., Giudici, M.L., Nipot, E. and Villa, B., 1988. Chemical control trials against rice blast in Italy. *Informatore Fitopatologic*, 38: 41-47.
- Rajan, C.P.D. 1987 Estimation of yield losses due to sheath blight of rice. *Indian Phytopath.* **40**: 174-177.
- Roy, A.K. 1993 Sheath blight of rice in India. Indian Phytopath 46: 97-205.
- Savary, S. and Mew, T.W. 1996. Analyzing crop losses due to *Rhizoctonia solani: rice sheath blight, a case study*. In: Sneh, B., Javaji-Hare, S., Neate, S., Dijst, G. (eds) *Rhizoctonia* species: taxonomy, molecular biology, ecology, pathology and disease control, Kluwer, Dordrecht, pp 237-244.
- Seebold, K.W., Datnof, J.L.E., Correa-Victoria, F.J., Kucharek, T.A, and Snyder, G.H, (2004). Effects of Silicon and fungicides on the control of leaf and neck blast in upland rice. *Plant Dis.* 88: 253-258.
- Singh, S., Mohan, C. and Pannu, P.P.S. 2014. Bioefficacy of different fungicides in managing blast of rice caused by Pyricularia grisea. *Plant Dis Res.* 29:16-20.
- Sood, G.K. and Kapoor, A.S. 1997. Efficacy of new fungicides in the management of rice blast. *Plant Dis. Res.* **12:**140-142.
- Sundravadana, S., Alice, D., Kuttalam, S., and Samiyappan, R. 2007. Azoxystrobin activity on *Rhizoctonia solani* and its efficacy against rice sheath blight. *Tunisian J. Plant Prot.* 2: 79-84.
- Swamy, H.N., Sannaulla and Dinesh, K.M. 2014. Screening some new fungicides against rice sheath blight disease. *Karnataka J. Agric. Sci.* 22:79-84.
- Tangdiabang, J. and Pakki, S. 2006 Penyakit blast (*Pyricularia grisea*) dan strategi pengendaliannya pada tanaman padi. *Pusat Penelitian Tanaman Pangan Departmen Pertanian* 7: 241-245.
- Varma, Y.C.K. and Santhakumari, P. 2012. Management of rice blast through new fungicidal formulation. *Indian Phytopath*, 65: 87-88.